Furcation perforation: current approaches and future perspectives

Manal Farea,¹ Adam Husein² and Cornelis H Pameijer³

¹ Manal Farea is a dentist with a BDS degree from Sana'a University, Sana'a, Yemen in 2003. She received her MSc degree in endodontics from Universiti Sains Malaysia (USM), Malaysia in 2010. In 2015, she completed her PhD degree at the USM in regenerative endodontics. Dr Manal was granted a scholarship from Sana'a University, Yemen in 2007 and a fellowship from USM in 2011.

² Professor Dr Adam Husein is a senior lecturer in the restorative unit (prosthodontics) and the dean of School of Dental Sciences, Universiti Sains Malaysia. He got his BDS from University of Adelaide, Australia in 1996. In 2004, he optained his graduate diploma in clinical dentistry, doctor in clinical dentistry and fellowship of the Royal Australasian College of Dental Surgeons (FRACDS) from the University of Adelaide.

³ Cornelis H Pameijer DMD MScD DSc PhD graduated from the University of Utrecht with a DDS in The Netherlands in 1967 and went on to further his studies at Boston University in the USA. He is currently professor emeritus at the University of Connecticut in Farmington, Connecticut, USA. He has lectured extensively worldwide and has published more than 300 publications in mostly peer-reviewed journals. During root canal treatment many procedural accidents may occur of which perforation of the root canal system plays a significant role. Perforation is defined by the American Association of Endodontics (AAE) Glossary of Endodontic Terms (2003) as a mechanical or pathological communication between the root canal system and the external tooth surface, which is caused by caries, resorption or iatrogenic factors. It has been identified as the second greatest cause of endodontic failure that accounts for 9.6% of all unsuccessful cases (Pitt Ford et al, 1995).

As a result of furcation perforation, destruction of the periodontal tissues may occur, which ultimately lead to loss of the tooth (Arens, Torabinejad, 1996; Tsesis, Fuss, 2006). The prognosis of the tooth depends upon several factors:

- 1. The severity of initial damage to the periodontal tissue
- 2. The location and size of perforations
- 3. The bacterial contamination
- 4. The sealing ability or cytotoxicity of the repair materials (Tsesis, Fuss, 2006; Sinai, 1977; Balla et al, 1991).

Even if a biocompatible material is used to treat a perforation, extensive injury may cause irreversible damage to the attachment apparatus at the furcation area (Sinai et al, 1989).

In large perforations, the complete sealing of the defect with a repair material is problematic and allows irritants to continuously penetrate into the furcation area (Balla et al, 1991). Perforations close to the gingival sulcus produce persistent inflammation and a down-growth of sulcular epithelium into the defect (Tsesis, Fuss, 2006). Sinai (1977) stated that coronally located perforations including furcal perforations were more serious than those in the middle and apical third of a canal. It is the objective of this review to collect and review the data that is available in the scientific literature and to reach a conclusion as to the best treatment options.

Methods

Retrieval of literature

An English-limited Medline search was performed of articles published from 2002 to 2015. The searched keywords included 'perforations and endodontics', 'furcation perforation', 'root canal and perforation', and 'perforation and mineral trioxide aggregate (MTA)'. Then, a hand search was done of the references of collected articles to determine if more papers relevant to the topic should be included.

Results

A total of 820 articles were found, which, in order of their related keywords, accounted for the following: perforations and endodontics: 285; furcation perforation: 92; root canal and perforation: 299; and perforation and mineral trioxide aggregate (MTA): 144.

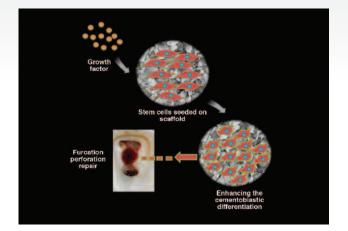


Figure 1: This illustration depicts a furcation perforation repair using stem cells, scaffold and growth factor. This method has the potential to open new avenues in furcation repair treatment in the foreseeable near future. This image relates to the text under 'future perspectives for the perforation repair' on page 40.

Perforation repair techniques and their prognosis

Surgical and non-surgical approaches have been utilised for periodontal tissue re-establishment at the perforation site. In both surgical and non-surgical approaches, two factors should be considered:

- 1. An appropriate material selection
- 2. The use of a matrix (Clauder, Shin, 2009).

The repair material should be selected based on the following criteria:

- Perforation site accessibility
- Biocompatibility (be nontoxic and noncarcinogenic)
- Ability to induce osteogenesis and cementogenesis
- Moisture control
- Easy handling

• Aesthetic considerations (Clauder, Shin, 2009; Bryan, Woollard, Mitchell, 1999; Yildirim et al, 2005; Samiee et al, 2010).

Matrix use

Controlling haemostasis and placement of the repair material in the perforation site without extrusion into surrounding periodontal structures are essential prerequisites for the success of a perforation repair. In order to achieve a fluidtight seal, haemostasis has to be controlled (Clauder, Shin, 2009). Delayed perforation repair can lead to extrusion of repair materials as a result of breakdown of the surrounding periodontium that is replaced by granulation tissue. Thus, in an attempt to avoid extrusion of the repair material, internal matrices such as calcium sulphate, hydroxyapatite, collagen, demineralised freeze-dried bone and Gelfoam have been used (Clauder, Shin, 2009; Roda, 2001; Bargholz, 2005).

The internal matrix concept was introduced by Lemon (1992) in order to adequately seal the furcation perforation and avoid extrusion of the material. He also recommended the use of hydroxyapatite as a matrix under amalgam. Calcium sulphate and calcium hydroxide prevented extrusion of composite resin when used as a furcal repair material (Imura et al, 1998). In 1999, Jantarat and colleagues demonstrated that amalgam placed with plaster of Paris as a matrix for furcal perforation repair improved its sealing ability. Hapset (65% non-resorbable hydroxyapatite and 35% plaster of Paris) and hydroxyapatite showed similar healing responses when used as internal matrices under amalgam (Rafter et al, 2002). Rafter et al (2002) further reported that there was marked extrusion of amalgam into the underlying bone with an associated severe inflammatory response when used alone without a matrix.

Although it has been reported that without using an internal matrix the optimal strength and excellent sealability of MTA was achieved in the presence of moisture (Arens, Torabinejad 1996; Holland et al, 2001; Torabinejad et al, 1994), conflicting results have been reported by some authors regarding the use of an internal matrix under MTA. In 2004, Kratchman suggested that the perforation site should be soaked with sodium hypochlorite after haemostasis had been achieved and that a physical barrier such as collagen or calcium sulfate must be used at the perforation site to prevent MTA from being packed into the bone.

According to Bargholz (2005), excellent clinical results were achieved when collagen matrix was used under MTA. A study by Al-Daafas and Al-Nazhan (2007) showed that calcium sulfate prevented extrusion of the repair material. However, an unfavourable inflammatory reaction - epithelial tissue migration into the defected perforation and the inability to induce bone regeneration - were detected. Thus, the authors concluded that using calcium sulphate as an internal matrix for MTA is not recommended. When used as an internal matrix for furcal perforation repair, calcium sulfate and Collaplug (Calcitek, Carlsbad, CA) did not improve the sealing ability nor reduce the incidence of MTA overextension. Therefore, the authors concluded that these two materials are not recommended as an internal matrix for MTA (Zou et al, 2008). Furthermore, calcium sulfate and hydroxyapatite did not improve the sealing ability of MTA when used as internal matrices for furcation perforation repair (Taneja, Kumari 2011).

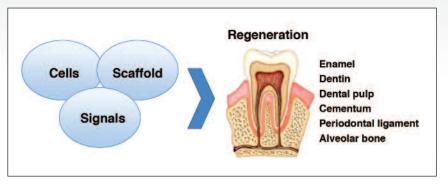


Figure 2: The three key elements of dental tissue engineering are stem cells, scaffolds and signals.

Materials used for furcation perforation repair

In an attempt to repair a furcation perforation, several materials such as amalgam, tricalcium phosphate (TCP), hydroxyapatite, gutta percha, calcium hydroxide, zinc oxideeugenol-based cement (IRM and Super-EBA), glass ionomer cement, composite resins, resin-glass ionomer hybrids, demineralised freeze-dried bone and MTA have been used over the years (Arens, Torabinejad, 1996; Balla et al, 1995; Bryan, Woollard, Mitchell, 1999, Yildirim et al, 2005; Salman et al, 1999). However, none fulfil all requisite qualifications for an ideal biomaterial.

Balla et al (1991) reported that no hard tissue was formed at the furcation perforation defect site when treated with either tri-calcium phosphate, hydroxyapatite, amalgam or calcium hydroxide (Life); instead, the defect site was occupied by epithelium and acute inflammatory cells (Balla et al, 1991). MTA is water-based cement that is derived from Portland cement (type I). It was introduced as a root-end filling material in the early 1990s (Torabinejad, Watson, Pitt Ford, 1993; Torabinejad, Chivian, 1999). It was subsequently determined that it was a suitable material for various clinical applications such as pulp capping, repair of furcal perforations as well as root-end closure (Sinai et al, 1989; Torabinejad et al, 1995). MTA promotes periradicular tissue regeneration (Pitt Ford et al, 1995; Yildirim et al, 2005; Holland et al, 2001; Zhu, Xia, Xia, 2003; Noetzel et al, 2006) and it differs from other materials by its ability to promote cementum regeneration, thus facilitating the regeneration of the periodontal apparatus (Pitt Ford et al, 1995; Arens, Torabinejad, 1996). Its biocompatibility nature is suggested by its ability to form hydroxyapatite when exposed to simulated body tissue fluid (Sarkar et al, 2005).

Two commercial forms of MTA are available; Proroot MTA (Dentsply Tulsa Dental), which is available in both gray or white form, of which the latter contains a lower amount of iron, and MTA-Angelus (Angelus) (Asgary et al, 2005). MTA-Angelus was introduced to address the long setting time from two hours for Proroot MTA to 10 minutes for MTA-Angelus. MTA-Angelus contains 80% Portland cement and 20% bismuth oxide, with no addition of calcium sulfate, while Proroot MTA is composed of 75% Portland cement, 20% bismuth oxide, and 5% calcium sulfate dehydrate (Hashem et al, 2008). The constituents of the Portland cement are minerals, amongst which the most important are dicalcium silicate, tricalcium silicate, tricalcium aluminate, tetracalcium ironaluminate and dehydrated calcium sulfate (Oliveira et al, 2007; Asgary et al, 2009a). The only significant difference between the dominant compounds of white and gray MTAs and associated Portland cements is bismuth oxide, which is present in MTAs (Asgary et al, 2009a; Asgary et al, 2004).

It has been reported that the sealing ability of MTA (Loma Linda University, Loma Linda, CA) was significantly better compared to amalgam in preventing leakage of Fusobacterium nucleatum through furcal perforations (Nakata, Bae, Baumgartner, 1998). When used to seal a large furcation perforation, Proroot MTA with/without internal matrix and MTA-Angelus with internal matrix showed the lowest dye absorbance compared to zinc oxide-eugenol cement (IRM) with/without internal matrix and MTA-Angelus without internal matrix. Additionally, the authors reported that IRM without internal matrix had the highest dye absorbance (Hashem, Hassanien, 2008). However, white and gray MTA (Dentsply Tulsa Dental) showed no significant differences in microleakage when used for furcal perforation repair (Ferris, Baumgartner, 2004; Hamad, Tordik, McClanahan, 2006). Furcal perforations have been repaired with Proroot gray MTA (Dentsply) and Geristore (Denmat). Geristore has been used as a root end filling material and in the restoration of subgingival surface defects such as root surface caries and iatrogenic perforations, surgical repair of root perforations

and as an adjunct in guided-tissue regeneration (GTR) (Mehrvarzfar et al, 2010). It also leaked significantly less than amalgam (Mehrvarzfar et al, 2010). In the aforementioned study, the authors reported that the sealing ability of MTA and Geristore was reduced when bioglass was used as a matrix underneath.

Sluyk, Moon and Hartwell (1998) assessed the effect of time and moisture on setting, retention and adaptability of MTA when used for furcal perforation repair. Findings showed that MTA adaptation to perforation walls increased in the presence of moisture. They further suggested that a moistened matrix can be used under MTA to prevent under- or overfilling of the material. Furthermore, Main et al (2004) indicated that MTA provided an effective seal for root perforations.

Yildirim et al (2005) investigated the histologic response to MTA and Super EBA (Bosworth Company) when used in furcation perforation repair in dogs. In their study, less inflammation and new cementum formation was observed with MTA compared to Super EBA, which demonstrated connective tissue repair without inflammation. Similar abilities to seal furcal perforations were observed for both Portland cement and MTA (De-Deus et al, 2006; Noetzel et al, 2006) evaluated histologically the inflammatory reactions and tissue responses to experimental tricalciun phosphate (TCP) and MTA when used as repair materials in furcation perforations in dogs. Results showed no significant differences between MTA and TCP in terms of bone reorganisation or deposition of fibrous connective tissue.

Thus, MTA is considered the gold standard and material of choice for perforation repair and has demonstrated good potential for clinical success. However, it has some disadvantages, including the inability to degrade to allow for replacement with natural tissues, low resistance to compression over the long-term, extended setting time, poor handling, and difficult insertion into cavities because of its granular consistency, while additional moisture is required to activate the cement setting, and lastly, the high cost, despite its widespread use (Torabinejad et al, 1995; Chng et al, 2005; Kogan et al, 2006; Coomaraswamy, Lumley, Hofmann, 2007; Parirokh, Torabinejad, 2010). Many dental materials have been demonstrated in the literature to exhibit cytotoxic effects during setting. Low cell numbers were demonstrated in vivo with freshly mixed MTA (pH=10.2) compared to preset MTA (pH=12.5) (Tronstad, Wennberg, 1980). However, histologically, no difference in bone and cementum regeneration was observed after periradicular surgery in dogs between fresh and preset Proroot MTA (Apaydin, Shabahang, Torabinejad, 2004).

In 2006, Asgary and colleagues introduced a new endodontic cement, a calcium-enriched mixture (CEM) cement. Major components of CEM cement powder are 51.75 wt.% calcium oxide, 9.53 wt.% sulfur trioxide, 8.49 wt.% phosphorous pentoxide, and 6.32 wt.% silicon dioxide; whereas the minor essential constituents are aluminium oxide > sodium oxide > magnesium oxide > chlorine. CEM cement has a similar pH but an increased flow compared to MTA. However, working time, film thickness and price are considerably less (Asgary et al, 2008a). Unlike MTA, mixed CEM cement releases calcium and phosphate ions and forms hydroxyapatite not only in simulated body tissue fluid but also in normal saline solution (Asgary et al, 2009a; Amini et al, 2009).

Although the chemical composition of CEM cement and MTA are different, they have similar clinical applications (Asgary et al, 2008b; Asgary et al, 2008c; Asgary et al, 2009b; Asgary, Ehsani, 2009c). Similar to MTA, CEM cement had low cytotoxic effects on different cell lines (Asgary et al, 2009d). However, it showed a better antibacterial effect comparable to calcium hydroxide (Asgary et al, 2008d). Similar sealing ability was demonstrated by both Proroot MTA and CEM when used to repair furcal perforation of primary molar teeth (Haghgoo et al, 2014).

Non-surgical approach

When a perforation repair is indicated, it is recommended to first attempt an intracoronal approach (non-surgical) to preserve the periodontium thus increasing the chances of success (Regan, Witherspoon, Foyle, 2005). Generally, perforations coronal to the crestal bone fall into the category of a non-surgical approach. The use of a surgical microscope operated at high magnification and with ample illumination allows for better management of perforation repairs (Kratchman, 2004; Daoudi, Saunders, 2002).

A surgical approach may complicate the treatment and lead to loss of periodontal attachment, chronic inflammation and furcal pocket formation (Arens, Torabinejad 1996). Experience has shown that buccally located perforations are easier to repair than lingual or proximal lesions. Lingual located perforations, especially in the mandible, should be treated non-surgically or orthodontically. If they are not responding to treatment, the tooth should be extracted (Regan et al, 2005). If a tooth can be extruded orthodontically to a point where the perforation reaches a supragingival level, repair of the defect will be greatly facilitated (Smidt, Lachish-Tandlich, Venezia, 2005). Whether clinically practical or not, one case of intentional reimplantation was reported after repair of the perforation was performed on

the extracted tooth (Poi et al, 1999).

In cases of large perforations, bleeding should be controlled first using sterile saline. Alternatively, calcium hydroxide, calcium sulphate, or collagen has been used (Clauder, Shin 2009). For bleeding control, non-specific intravascular clotting agents should be avoided as they may lead to alveolar bone damage and delay in healing (Lemon, Steele, Jeansonne, 1993). In cases of perforations that are infected or perforation sites that need further enlargement and cleaning, burs or ultrasonic tips may be used. However, ultrasonic tips are preferable as they are gentler to the adjacent periodontium tissues (Pitt Ford et al, 1995; Arens, Torabinejad, 1996; Clauder, Shin, 2009). For cleaning of infected perforations, 2.5% sodium hypochlorite has been used (Arens, Torabinejad, 1996), however, sterile saline is indicated in large perforations (Clauder, Shin, 2009). To avoid blockage of the canals with repair material, gutta percha points, paper points, cotton pellets or an easily removable material (such as Cavit) should be placed over the canal orifices (Clauder, Shin, 2009).

A resin-bonded material such as Geristore (Denmat) is recommended to restore subgingival defects (Clauder, Shin, 2009), which also serves as an adjunct to GTR (Abitbol et al, 1996; Behnia, Strassler, Campbell, 2000). It is less sensitive to moisture than conventional glass ionomer cement while a drier environment improved the results (Cho, Kopel, White, 1995). Adhesive materials can be used in supracrestal perforations, whereas MTA is preferable in subcrestal perforations (Clauder, Shin, 2009). If a perforation defect involves bone destruction (intraosseus defect), a barrier is needed to facilitate controlled placement of the repair material. This is not necessaary if the defect does not include an intraosseus defect (Clauder, Shin, 2009). If MTA is used a moist cotton pellet should cover the material to allow setting of the material. After perforation repair the final restoration can be placed either after one day or one week. Once repair has been achieved the root canal(s) can be cleaned, shaped and filled (Pitt Ford et al, 1995; Arens, Torabinejad, 1996).

If a perforation is present in the middle third of the root, the canal(s) should be prepared first before closing the defect to avoid blocking the canal. With the aid of an operating microscope, obturation of the canal apical to the defect should be done first, followed by filling the remainder of the canal and the perforation site with MTA (Clauder, Shin, 2009). Alternatively, the root space beyond the perforation can be maintained by means of a file or gutta percha cone. In case a file is used, it should be loosened after finishing the repair procedure to allow easy removal before the MTA is fully set (Clauder, Shin, 2009). The other option is to use a gutta percha point and soften it with heat to the dentinal wall opposing the perforation. MTA is then placed at the defect site (Clauder, Shin, 2009). Perforations at the apical one-third are quite challenging and difficult to manage. Successful treatment cannot always be achieved for all cases necessitating apical surgery or extraction of the tooth to remedy the problem (Clauder, Shin, 2009).

Surgical approach

Surgical intervention (external approach) is indicated in areas that are not accessible by non-surgical means alone, cases that have not responded to non-surgical treatment or in repairing a perforating resorption (Regan et al, 2005). The surgical approach is performed by reflecting a flap at the perforation site followed by cleaning and preparing the perforated area and finally packing the repair material (Alhadainy, 1994).

During the surgical repair procedures, cortical bone damage is involved, which may result in reduced success of the corrective surgical procedure. Thus, a GTR technique has been recommended for successful treatment outcomes by using either non-resorbable or resorbable membranes as a barrier (Duggins et al, 1994; Barkhordar, Javid 2000; Rankow, Krasner, 1996; Dean et al, 1997; Leder et al, 1997). This barrier guides selected cells to populate at the perforation defect, ie, placing the barrier between the gingival tissue and the perforation defect will facilitate the repopulation of the defect by periodontal ligament cells and other osteogenic cells and prevents the colonisation by gingival cells (Linde et al, 1993; Sandberg, Dahlin, Linde, 1993). A resorbable membrane is generally preferable, as it does not need a second surgical procedure to remove it. However, in some cases, titanium-tented membrane or a supporting graft material is needed to prevent collapsing the membrane into the defect (Abitbol et al, 1996).

Cementum regeneration and role in the periodontium reconstruction

Cementum formation is very essential in the furcation perforation repair process (Pitt Ford et al, 1995; Clauder, Shin, 2009; Samiee et al, 2010; Zairi et al, 2012). Pitt Ford and colleagues (1995) evaluated the histologic response to experimentally induced furcation perforations in dog mandibular premolars repaired by either MTA or amalgam and found that most of the MTA samples showed no inflammation and cementum deposition, whereas with the use of amalgam, moderate to severe inflammation with no cementum deposition was present.

Healing after intentional perforations in dogs' teeth was evaluated after repair with either MTA or Sealapex (Kerr) (Holland et al, 2001). Most samples sealed with MTA showed new cementum deposition and an absence of inflammation. In 2010, Samiee and colleagues reported that cementum-like hard tissue was formed using either MTA or CEM cement in the furcation perforation in dogs in the presence of a mild inflammatory response. The authors concluded that both materials showed a similar favourable biological response in furcation perforation repair.

Zairi et al (2012) compared the inflammatory reactions and tissue response of furcal perforations in dogs' teeth to growth factors, TGF β 1, basic fibroblast growth factor (bFGF), osteogenic protein-1 (OP-1) and IGF-1, with MTA or IRM as controls. The authors reported that a clear stimulatory effect on cementum formation and inhibition of collagen capsule formation was exerted by the growth factors. However, MTA exhibited better results than the growth factors. Based on that, the authors suggested a further study

20

comparing the effects of application of growth factor mixture with MTA and MTA alone on tissue healing and regeneration.

In a case report, Bains et al (2012) used tissue engineering principles for the furcation perforation repair of the pulpal floor of the right mandibular first molar of 39-yearold male patient using MTA and platelet-rich fibrin (PRF). The authors reported that this combination was able to repair the perforation defect and regenerate the lost periodontium in the furcation area effectively. A case report (Eghbal, Fazlyab, Asgary, 2014) was published describing the nonsurgical endodontic management of an extensive perforation of the floor of the pulp chamber in a first mandibular molar of a 28-year-old Caucasian female using CEM cement. The authors reported that CEM was able to induce hard tissue formation, ie bone and cementum.

Cellular tissue engineering approach for cementum regeneration

A proposed therapeutic approach was reported by the

www.ids-cologne.de

38th International Dental Show **Cologne, 12–16 March 2019** Trade Dealer Day: 12 March 2019

LEADING DENTAL BUSINESS SUMMIT

Admission ticket = public transport ticket Free local travel to and from IDS

South Africa

Southern African-German Chamber of Commerce and Industry No 47, Oxford Road, Forest Town 2193, Johannesburg P.O.Box 87078, Houghton 2041 Tel. +27 11 4862775, Fax +27 (0) 86 501 1888 dvanjaarsveldt@germanchamber.co.za







removal of autologous cells from the patient's periodontal ligament (PDL), culturing of the cells in vitro, which were then placed back onto the exposed root coated with chemo attracting factors, subsequently covering the area with an artificial basement membrane (Terranova, 1990). However, it is unknown whether this method produced the desired effect. Lekic and colleagues (2005) reported that rat periodontal and bone marrow cells were able to differentiate into periodontal ligament fibroblasts, osteoblasts and cementoblasts when transplanted into periodontal wounds in rats, thus contributing to periodontal regeneration.

Regeneration of cementum, PDL and alveolar bone have been observed using auto-transplantation of bone marrow derived mesenchymal stem cells (B/MMSCs) (Kawaguchi et al, 2004) or periodontal ligament cell sheet (Akizuki et al, 2005) into periodontal osseous defects in dogs. However, the principle disadvantage of cell sheets is their delicate structure and difficult handling during surgery (Li, Jin, 2015). Furthermore, the harvest of bone marrow (BM) is a highly invasive and a painful procedure for the donor. Moreover, it has been reported that the number, proliferation and differentiation potential of B/MMSCs decline with increasing age (Kern et al, 2006).

It has been reported that cementoblast-biodegradable poly(lactic-co-glycolic acid) (PLGA) polymer sponge-treated defects showed complete bone bridging and PDL formation, whereas minimal evidence of osteogenesis was exhibited by follicle cell-treated defects along the root surface of athymic rats (Zhao et al, 2004). Periodontal ligament stem cells (PDLSCs) have the ability to differentiate into cementoblast and osteoblast (Isaka et al, 2001; Seo et al, 2004) and have shown potential therapeutic applications in periodontium regeneration. However, the very low number of these cells residing in the PDL is indicative of the difficulty acquiring a sufficient number for regenerative treatment remains and is an issue that remains unresolved (Maeda et al, 2011). Primary cultures of PDLSCs yielded small cell numbers, therefore before application, PDLSCs must proliferate at least 12 population doublings (Zhu, Liang, 2015). Additionally, it has been found that the proliferation and migration ability and differentiation potential of PDLSCs decreased with increasing age (Zhu, Liang, 2015).

Apical tooth germ cells conditioned medium were able to provide the cementogenic microenvironment and induced the cementoblastic differentiation of PDLSCs (Yang et al, 2009). Hertwig's epithelial root sheath (HERS) cells, or their secreted products, were able to induce PDL cells differentiation along the cementoblastic lineage in vitro (Zeichner-David et al, 2003). Several in vivo studies have also shown the potential capability of PDLSCs to form cementum and PDL-like tissues (Yang et al, 2009; Liu et al, 2008; Feng et al, 2010; Park, Jeon, Choung, 2011).

Regenerative therapy

Tissue engineering is an interdisciplinary field that applies the principles of engineering and life sciences toward the development of biological substitutes that restore, maintain, or improve tissue function or a whole organ (Langer, Vacanti, 1993). Tissue engineering aims to stimulate the body either to regenerate tissue on its own or to grow tissue outside the body, which can then be implanted as natural tissue (Nadig, 2009).

Triad components

Regenerative endodontics can be defined as biologically based procedures designed to replace damaged structures, including dentine and root structures, as well as cells of the pulp-dentine complex (Murray, Garcia-Godoy, Hargreaves, 2007). This approach consists of the following interactive triad: 1) an appropriate cell source; 2) a supportive matrix (scaffold); and 3) inductive biological factors or signals (Figure 1). To create regenerative therapies, these disciplines are often combined rather than used individually (Murray, Garcia-Godoy, Hargreaves, 2007).

Future perspectives for the perforation repair

Reconstruction of the lost attachment via regeneration of the periodontium components, such as cementum, PDL and bone, is essential in the repair of perforated areas. Replacement of the lost cementum (cementogenesis) is very critical and enhances the reattachment of the fibres of the periodontal ligament. Several studies have been published that demonstrate the ability of different materials to repair furcation perforations, albeit with variable success rates.

However, during recent years, there has been a paradigm shift from conventional to regenerative endodontic therapy and repair of the periodontium is not an exception. To date, to the best of our knowledge, no studies have been published in the literature reporting on the effect of the triad application (stem cells, scaffold and growth factor) for furcal perforation repair and the response of surrounding tissues (cementum, PDL and alveolar bone). We propose a stem cellbased tissue engineering approach for furcation perforation repair through enhancing of stem cell differentiation along the cementoblastic lineage in association with scaffold and growth factor. The suggested biomimetic approach is illustrated in Figure 2. This will have the potential to open a new era and strategy in endodontic and periodontal tissue engineering therapies.

Conclusions

Perforation of the pulp chamber floor of multi-rooted teeth constitutes a perplexing and frustrating problem. It is a major cause of endodontic treatment failure. A furcation perforation has to be regarded as an endodontic and periodontal problem. The inflammatory response in the periodontium, leading to irreversible loss of periodontal attachment in the area, can result in loss of the tooth if the perforation is not successfully repaired. To re-establish the periodontal tissue in the perforation site, surgical and non-surgical techniques have been utilised.

For furcation perforation repair, several materials have been used with varying results. However, the stem cell-based tissue engineering approach is very promising and is suitable for furcation perforation repair. This approach has the potential to revolutionise the practice of regenerative

ncisal

endodontics in the future and may therefore save many teeth that would otherwise have to be extracted due to a poor to hopeless prognosis.

Moreover, it will help and assist in designing regenerative therapies based on sound biological principles, which can be applied in both endodontic and periodontal specialties.

Acknowledgements

This study was financially supported by the Universiti Sains Malaysia Research University Grant 1001/PPSP/813058, PRGS (1001/PPSG/8146005) and short-term grants (304/PPSG/ 61312012 and 304/PPSG/61312018) from the School of Dental Sciences, Universiti Sains Malaysia.

References

A full list of references is available from the Publisher

Reprinted with permission by Endodontic Practice August 2017

- Infection Control Specialist
- Dental Assistant Training
- Specialised Consulting
- Marketing and Practice Management



Melanie Savvides has worked in the Dental Industry for the last 32 years and was the MD of one of the largest Dental supply companies in South Africa. She has travelled around the world through dentistry, attending numerous courses, workshops and events.

Melanie is passionate about Dentistry in South Africa and would like to share her experience with you.



Abitbol T, Santi E, Scherer W, Palat M (1996) Using a resin-ionomer in guided tissue regenerative procedures: technique and application-case reports. Periodontal Clin Investig 18:17–21

Akizuki T, Oda S, Komaki M, Tsuchioka H, Kawakatsu N, Kikuchi A, Yamato MJ, Okano T, Ishikawa I (2005) Application of periodontal ligament cell sheet for periodontal regeneration: a pilot study in beagle dogs. J Periodontal Res 40:245–251

Al-Daafas A, Al-Nazhan S (2007) Histological evaluation of contaminated furcal perforation in dogs' teeth repaired by MTA with or without internal matrix. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 103:92–99

Alhadainy HA (1994) Root perforations. A review of literature. Oral Surg Oral Med Oral Pathol 78:368–374

American Association of Endodontists (2003) Glossary of Endodontic Terms. Chicago, Illinois, USA

Amini Ghazvini S, Abdo Tabrizi M, Kobarfard F, Akbarzadeh Baghban AR, Asgary S (2009) Ion release and pH of a new endodontic cement, MTA and Portland cement. Iranian Endod J 4:74–78

Apaydin ES, Shabahang S, Torabinejad M (2004) Hard-tissue healing after application of fresh or set MTA as rootend-filling material. J Endod 30:21-24

Arens DE, Torabinejad M (1996) Repair of furcal perforations with mineral trioxide aggregate: two case reports. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 82:84–88

Asgary S, Akbari Kamrani F (2008d) Antibacterial effects of five root canal sealing materials. J Oral Scie 50:469–474

Asgary S, Eghbal M J, Parirokh M, Torabzadeh H (2006) Sealing ability of three commercial mineral trioxide aggregates and an experimental rootend filling material. Int Endod J 1:101–105

Asgary S, Eghbal MJ, Parirokh M (2008a) Sealing ability of a novel endodontic cement as a root-end filling material. J Biomed Mater Res A 87:706–709

Asgary S, Eghbal MJ, Parirokh M, Brink F (2005) Chemical differences between white and gray mineral trioxide aggregate. J Endod 31:101–103

Asgary S, Eghbal MJ, Parirokh M, Ghanavati F, Rahimi H (2008c) A comparative study of histological response towards different pulp capping materials and a novel experimental cement. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 106:609–614

Asgary S, Eghbal MJ, Parirokh M, Ghoddusi J (2009b) Effect of two storage solutions on surface topography of two rootend fillings. Aust Endod J 35:147–52

Asgary S, Eghbal MJ, Parirokh M, Ghoddusi J, Kheirieh S, Brink F (2009a) Comparison of mineral trioxide aggregate's composition with Portland cements and a new endodontic cement. J Endod 35:243–250

Asgary S, Ehsani S (2009c) Permanent molar pulpotomy with a new endodontic cement: a case series. J Conserv Dent 12:31–36 $\,$

Asgary S, Moosavi SH, Yadegari Z, Shahriari S (2009d) Cytotoxic effect of MTA and New Endodontic Cement in human gingival fibroblast cells: a SEM evaluation. N Y State Dent.] 78:51–4

Asgary S, Parirokh M, Eghbal MJ, Brink F (2004) A comparative study of white mineral trioxide aggregate and white Portland cements using X-ray microanalysis. Aust Endod J 30:89–92 (Abstract)

Asgary S, Shahabi S, Jafarzadeh T, Amini S, Kheirieh S (2008b) The properties of a new endodontic material. J Endod 34:990–993.

Bains R, Bains V, Loomba K, Verma K, Nasir A (2012) Management of pulpal floor perforation and grade π furcation involvement using mineral trioxide aggragate and platelet rich fibrin: A clinical report. Contemp Clin Dent 3:223–227

Balla R, LoMonaco CJ, Skribner J, Lin LM (1991) Histological study of furcation perforations treated with tricalcium phosphate, hydroxylapatite, amalgam, and life. J Endod 17:234–238

Bargholz C (2005) Perforation repair with mineral trioxide aggregate: a modified matrix concept. Int Endod J 38:59–69

Barkhordar RA, Javid B (2000) Treatment of endodontic perforations by

guided tissue regeneration. Gen Dent 48:422-426

Behnia A, Strassler HE, Campbell R (2000) Repairing iatrogenic root perforations. J Am Dent Assoc 131:196–201

Bryan EB, Woollard G, Mitchell WC (1999) Nonsurgical repair of furcal perforations: a literature review. Gen Dent 47:274–278

Chng HK, Islam I, Yap AU, Tong YW, Koh ET (2005) Properties of a new root-end filling material. J Endod 31:665–668

Cho E, Kopel H, White SN (1995) Moisture susceptibility of resin-modified glass-ionomer materials. Quintessence Int 26:351–8

Clauder T, Shin SU (2009) Repair of perforations with MTA: clinical applications and mechanisms of action. Endod Topics 15:32–55

Coomaraswamy KS, Lumley PJ, Hofmann MP (2007) Effect of bismuth oxide radioopacifier content on the material properties of an endodontic Portland cement based (MTA-like) system. J Endod 33:295-8

Daoudi MF, Saunders WP (2002) In vitro evaluation of furcal perforation repair using mineral trioxide aggregate or resin modified glass lonomer cement with and without the use of the operating microscope. J Endod 28:512–515

De-Deus G, Petruccelli V, Gurgel-Filho E, Coutinho-Filho T (2006) MTA versus Portland cement as repair material for furcal perforations: a laboratory study using a polymicrobial leakage model. Int Endod J 39:293–8

Dean JW, Lenox RA, Lucas FL, Culley WL, Himel VT (1997) Evaluation of a combined surgical repair and guided tissue regeneration technique to treat recent root canal perforations. J Endod 23:525–532

Duggins LD, Clay JR, Himel VT, Dean JW (1994) A combined endodontic retrofill and periodontal guided tissue regeneration technique for the repair of molar endodontic furcation perforations: report of a case. Quintessence Int 25:109–114

Eghbal MJ, Fazlyab M, Asgary S (2014) Repair of an extensive furcation perforation with CEM cement: a case study. Iran Endod J 9:79–82

Feng F, Akiyama K, Liu Y, Yamaza T, Wang TM, Chen JH, Wang BB, Huang GT, Wang S, Shi S (2010) Utility of PDL progenitors for in vivo tissue regeneration: a report of 3 cases. Oral Dis 16:20–28

Ferris DM, Baumgartner JC (2004) Perforation repair comparing two types of mineral trioxide aggregate. J Endod 30:422–424

Haghgoo R, Niyakan M, Nazari Moghaddam K, Asgary S, Mostafaloo N (2014) An in vitro comparison of furcal perforation repaired with Pro-root MTA and new endodontic cement in primary molar teeth – a microleakage study. J Dent Shiraz Univ Med Sci 15:28-32

Hamad HA, Tordik PA, McClanahan SB (2006) Furcation perforation repair comparing gray and white MTA: a dye extraction study. J Endod 32:337–340

Hashem AA, Hassanien EE (2008) ProRoot MTA, MTA-Angelus and IRM used to repair large furcation perforations: sealability study. J Endod 34:59-61

Holland R, Filho JA, de Souza V, Nery MJ, Bernabe PF, Junior ED (2001) Mineral trioxide aggregate repair of lateral root perforations. J Endod 27:281-4

Imura N, Otani SM, Hata G, Toda T, Zuolo ML (1998) Sealing ability of composite resin placed over calcium hydroxide and calcium sulphate plugs in the repair of furcation perforations in mandibular molars: a study in vitro. Int Endod J 31:79–84

Isaka J, Ohazama A, Kobayashi M, Nagashima C, Takiguchi T, Kawasaki H, Tachikawa T, Hasegawa K (2001) Participation of periodontal ligament cells with regeneration of alveolar bone. J Periodontol 72:314–23

Jantarat J, Dashper SG, Messer HH (1999) Effect of matrix placement on furcation perforation repair. J Endod 25:192–196

Kawaguchi H, Hirachi A, Hasegawa N, Iwata T, Hamaguchi H, Shiba H, Takata T, Kato Y, Kurihara H (2004) Enhancement of periodontal tissue regeneration by transplantation of bone marrow mesenchymal stem cells. J Periodontol 75: 1281–1287

Kern S, Eichler H, Stoeve J, Kluter H, Bieback K (2006) Comparative analysis of mesenchymal stem cells from bone marrow, umbilical cord blood, or adipose tissue. Stem Cells 24:1294–301

Kogan P, He J, Glickman GN, Watanabe I (2006) The effects of various additives on setting properties of MTA. J Endod 32: 569-72

Kratchman SI (2004) Perforation repair and one-step apexification procedures. Dent Clin North Am 48:291–307

Langer R, Vacanti JP (1993) Tissue engineering. Science 260: 920–926 Leder AJ, Simon BI, Deasy M, Fenesy KE, Dunn S (1997) Histological, clinical, and digital subtraction radiographic evaluation of repair of periodontal defects resulting from mechanical perforation of the chamber floor using ePTFE membranes. Periodontal Clin Invest 19:9–15

Lekic PC, Nayak BN, Al-Sanea R, Tenenbaum H, Ganss B, McCulloch C (2005) Cell transplantation in wounded mixed connective tissues. Anat Rec A Discov Mol Cell Evol Biol 287:1256–1263

Lemon RR (1992) Nonsurgical repair of furcation defects (Internal matrix concept). Dent Clin North Am 36:439–457

Lemon RR, Steele PJ, Jeansonne BG (1993) Ferric sulfate hemostasis: effect on osseous wound healing. Left in situ for maximum exposure. J Endod 19:170–173

Li B, Jin Y (2015) Periodontal tissue engineering: Current approaches and future therapies. In: Stem Cell Biology and Tissue Engineering in Dental Sciences, Elsevier, UK 417–480

Linde A, Alberius P, Dahlin C, Bjurstam K, Sundin Y (1993) Osteopromotion: a soft+tissue exclusion principle using a membrane for bone healing and bone neogenesis. J Periodontol 64:1116–1128

Liu Y, Zheng Y, Ding G, Fang D, Zhang, C, Bartold PM, Gronthos S, Shi S, Wang S (2008). Periodontal ligament stem cell-mediated treatment for periodontitis in miniature swine. Stem Cells 26:1065–1073

Maeda H, Tomokiyo A, Fujii S, Wada N, Akamine A (2011) Promise of periodontal ligament stem cells in regeneration of periodontium. Stem Cell Res Ther 2:33

Main C, Mirzayan N, Shabahang S, Torabinejad M (2004) Repair of root perforations using mineral trioxide aggregate: a long-term study. J Endod 30:80–3

Mehrvarzfar P, Dahi-Taleghani A, Saghiri MA, Karamifar K, Shababi B, Behnia A (2010) The comparison of MTA, Geristore® and Amalgam with or without Bioglass as a matrix in sealing the furcal perforations (in vitro study). Saudi Dent J 22:119–24

Murray PE, Garcia-Godoy F, Hargreaves KM (2007) Regenerative endodontics: a review of current status and a call for action. J Endod 33:377–90

Nadig RR (2009) Stem cell therapy – Hype or hope? A review J Conserv Dent 12: $131{-}138$

Nakata T, Bae K, Baumgartner J (1998) Perforation repair comparing mineral trioxide aggregate and amalgam using an anaerobic bacterial leakage model. J Endod 24:184–6

Noetzel J, Ozer K, Reisshauer BH, Anil A, Rossler A, Neumann K, Kielbassa AM (2006) Tissue responses to an experimental calcium phosphate cement and mineral trioxide aggregate as materials for furcation perforation repair: a histological study in dogs. Clin Oral Invest 10:77–83

Oliveira MG, Xavier CB, Demarco FF, Pinheiro AL, Costa AT, Pozza DH (2007) Comparative chemical study of MTA and Portland cements. Braz Dent J $18{:}3{-}7$

Parirokh M, Torabinejad M (2010) Mineral trioxide aggregate: a comprehensive literature review-Part III: Clinical applications, drawbacks, and mechanism of action. J Endod 36:400-13

Park, JY, Jeon, SH, Choung PH (2011) Efficacy of periodontal stem cell transplantation in the treatment of advanced periodontitis. Cell Transplant 20: 271–85

Pitt Ford TR, Torabinejad M, McKendry DJ, Hong CU, Kariyawasam SP (1995) Use of mineral trioxide aggregate for repair of furcal perforations. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 79:756–763

Poi WR, Sonoda CK, Salineiro SL, Martin SC (1999) Treatment of root perforation by intentional reimplantation: a case report. Endod Dent Traumatol 15:132–134 Rafter M, Baker M, Alves M, Daniel J, Remeikis N (2002) Evaluation of healing with use of an internal matrix to repair furcation perforations. Int Endod J 35:775-783

Rankow HJ, Krasner PR (1996) Endodontic applications of guided tissue regeneration in endodontic surgery. J Endod 22:34–43

Regan JD, Witherspoon DE, Foyle DM (2005) Surgical repair of root and tooth perforations. Endod Topics 11:152–178

Roda RS (2001) Root perforation repair: surgical and nonsurgical management. Pract Proced Aesthet Dent 13:467–472

Salman MA, Quinn F, Dermody J, Hussey D, Colaffey N (1999) Histological evaluation of repair using a bioresorbable membrane beneath a resin-modified glass ionomer after mechanical furcation perforation in dogs teeth. J Endod 25:181–186

Samiee M, Eghbal MJ, Parirokh M, Abbas FM, Asgary S (2010) Repair of furcal perforation using a new endodontic cement. Clin Oral Investig 14:653–8

Sandberg E, Dahlin C, Linde A (1993) Bone regeneration by the osteopromotion technique using bioabsorbable membranes: an experimental study in rats. J Oral Maxillofac Surg 51:1106–1114

Sarkar NK, Caicedo R, Ritwik P, Moiseyeva R, Kawashima I (2005) Physicochemical basis of the biologic properties of mineral trioxide aggregate. J Endod 31:97–100

Seo BM, Miura M, Gronthos S, Bartold PM, Batouli S, Brahim J, Young M, Robey PG, Wang CY, Shi S (2004) Investigation of multipotent postnatal stem cells from human periodontal ligament. Lancet 364:149–55

Sinai I (1977) Endodontic perforations: their prognosis and treatment. J Am Dent Assoc $95{:}90{-}95$

Sinai IH, Romea DJ, Glassman G, Morse DR, Fantasia J, Furst ML (1989) An evaluation of tricalcium phosphate as a treatment for endodontic perforations. J Endod 15:399–403

Sluyk SR, Moon PC, Hartwell GR (1998) Evaluation of setting properties and retention characteristics of mineral trioxide aggregate when used as a furcation perforation repair material. J Endod 24:768–71

Smidt A, Lachish-Tandlich M, Venezia E (2005) Orthodontic extrusion of an extensively broken down anterior tooth: a clinical report. Quintessence Int 36:89–95

Taneja S, Kumari M (2011) Effect of internal matrices of hydroxyapatite and calcium sulfate on the sealing ability of mineral trioxide aggregate and light cured glass ionomer cement. J Conserv Dent 14:6-9

Terranova VP (1990) Periodontal and bone regeneration factor, materials and methods. International patent # VVO 90/ 100017

Torabinejad M, Chivian N (1999) Clinical applications of mineral trioxide aggregate. J Endod 25:197–205

Torabinejad M, Higa RK, McKendry DJ, Pitt Ford TR (1994) Dye leakage of four root end filling materials: effects of blood contamination. J Endod 20:159–163

Torabinejad M, Hong CU, Lee SJ, Monsef M, Pitt Ford TR (1995) Investigation of mineral trioxide aggregate for root-end filling in dogs. J Endod 21:603–608

Torabinejad M, Hong CU, McDonald F, Pitt Ford TR (1995) Physical and chemical properties of a new rootend filling material. J Endod 21:349-53

Torabinejad M, Watson TF, Pitt Ford TR (1993) Sealing ability of a mineral trioxide aggregate when used as a root end filling material. J Endod 19:591–5

Tronstad L, Wennberg A (1980) In vitro assessment of the toxicity of filling materials. Int Endod J 13:131–138

Tsesis I, Fuss Z (2006) Diagnosis and treatment of accidental root perforations. Endod Topics $13{:}95{-}107$

Yang ZH, Zhang XJ, Dang NN, Ma ZF, Xu L, Wu JJ, Sun YJ, Duan YZ, Lin Z, Jin Y (2009) Apical tooth germ cell-conditioned medium enhances the differentiation of periodontal ligament stem cells into cementum/periodontal ligament-like tissues. J Periodontal Res 44:199–210

Yildirim T, Gencoglu N, Firat I, Perk C, Guzel O (2005) Histologic study of